

Horizontal distribution of ectomycorrhizal infection in *Dipterocarpus turbinatus* plantations of Bangladesh

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Abstract: Garjan (*Dipterocarpus turbinatus* Gaertn. F) is a highly ectomycorrhizal tree species growing in hilly areas of Bangladesh. The horizontal distributions of ectomycorrhizas (ECM) in different distances (1, 2, 3 & 4m) from the tree base were determined in 5, 10 and 20-year-old *D. turbinatus* plantations of Chittagong University Campus (CUC) in 2003. The ECM infection (%) in roots was counted at three hill positions (top, mid and bottom) for each of the plantations. Samples of 1000 cm³ rhizosphere soil were collected from underneath the trees at different horizontal distances. The percentage of infection at different distances and hill elevations varied considerably. In 5 and 10-year-old plantations, the occurrence of infection (%) was rapidly declined with increasing distances, while in 20-year-old plantation, the infection increased sharply with increasing distances from the tree base. The highest infection (81.33%) was found at 4 m distance from the tree at bottom hill in 20-year-old plantation and the lowest (55.33%) at the same distance at the top of the hill in 5-year-old plantation.

Keywords: *Dipterocarpus turbinatus* plantation; Garjan; Ectomycorrhizas; Horizontal Distribution; Infection; Bangladesh

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Introduction

Dipterocarpus turbinatus, locally known as 'Telly Garjan', is a commercially important timber species naturally grown in Hill Forests of Bangladesh. It also grows in tropical evergreen and semi evergreen forests of the Andamans, greater part of Myanmer, Chittagong Hill Tracts and Cox's Bazar. The wood is good for making lorry bodies, boat building, railway sleepers, transmission poles and other construction purposes (Das 1980). Mycorrhizas are the symbiotic association between specialized root inhabiting fungi and the roots of living plants (Lee 1998). They play a significant role in plant nutrition, growth improvement, successful afforestation, reforestation, bio-control of pathogens and land reclamation programmes (Marx 1977; Rawat *et al.* 2003). All members of the Dipterocarpaceae so far examined are ectomycorrhizas (Singh 1966; Bakshi 1974; Hong 1979; de Alwis and Abeyanake 1980; Ashton 1982; Becker 1983; Alexander and Höglberg 1986; Smits 1994; Aniwat 1987; Hadi and Santoso 1988 and Hadi *et al.* 1991). They increase the tolerance of trees against drought, high soil temperatures, organic and inorganic toxins and extreme soil acidity (Lee 1998). Although a considerable amount of work was done on ECM in different parts of the world (Becker 1983; Hadi and Santoso 1988; Yasman 1993; Zarate *et al.* 1993), little has been done in Bangladesh (Shayesta and Choudhury 1985; Rahman and Mridha 2004). The present study was undertaken to explore the horizontal distribution of ECMs in 5, 10 and 20-year-old *D. turbinatus* plantations at three hill positions (top, mid and bottom).

Materials and methods

Study site

The study was carried out in 5, 10 and 20-year-old *D. turbinatus* plantations on hills of Chittagong University Campus (CUC), Bangladesh. The area lies between about 22°27'30" and 22°29'0" North latitudes and 91°46'30" and 91°47'45" East longitudes and covers about 1,271 acres of land approximately (Anon 1989). The hills are low to medium high and slope ranges from gentle to steep (Anon, 1979). Soils are yellowish brown to yellowish red loamy sand and weak to strong blocky. The sandy loam soil had moisture content around 25 percent and pH 5.6.

Assessment of ECM infection at various distances from a tree

In April 2003, ECM root samples were collected from points 1, 2, 3 and 4m apart from the base of a tree at three slope positions (top, mid and bottom) of each plantation. Three soil samples for each distance were randomly collected. To determine the horizontal distribution of ECMs, root samples along with rhizosphere soils (1000 cm³) were taken. Each collected sample was transferred into a separate plastic bag and marked. All samples were brought to laboratory to examine ECM association in the root tips. Fine feeder roots i.e., short roots were washed over 0.2 mm mesh-size sieve using a gentle flow of tap water. Then the short roots were transferred to the petri dishes and observed under compound microscope. The presence of ECMs in root tips was determined with the help of laboratory manual (Ingleby *et al.*, 1990). From each sample 100 fine roots were examined. When there were less than 100 fine roots all of them were examined and recorded. Then the percentage of ECM infection was calculated by using the following formula:

$$\text{ECM infection (\%)} = \frac{\text{Total number of infected root tips}}{\text{Total number of root tips studied}} \times 100$$

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Results

Distances from the tree base influence the formation of ECM root infection. The observations of such infection in plantations of three different ages (20, 10 & 5 years) are presented in Figures 1, 2 & 3. The occurrence of ECM infection (%) increased with distance from tree base in 20-year-old plantation (Figure 1). The highest infection (81.33%) was found in 4 m distance from the tree at bottom hill and the lowest (61.67%) at 1m distance at top hill position. It has been found in Figure 2 that the infection (%) was rapidly climbed between 1m and 2m distances from trees in 10-year-old plantation and then decreased gradually up to 4 m one. Maximum infection (73.33%) was recorded in 2m distance from the tree at the bottom and minimum (61.00%) was in 1m distance at top hill (Figure 2). In case of 5-year-old plantation, the percentage of ECM infection was gradually increased up to 2 m distance and then it declined sharply. The highest infection (74.33%) was found in 2 m distance at the bottom while the lowest (55.33%) –in 1 m at the top of the hill (Figure 3).

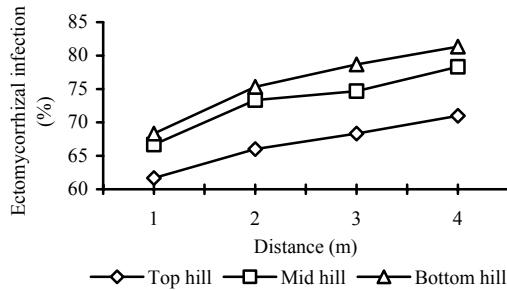


Fig. 1 ECM infection (%) in soil at 1, 2, 3 and 4m distances from the tree at three hill positions of 20-year-old *D. turbinatus* plantation

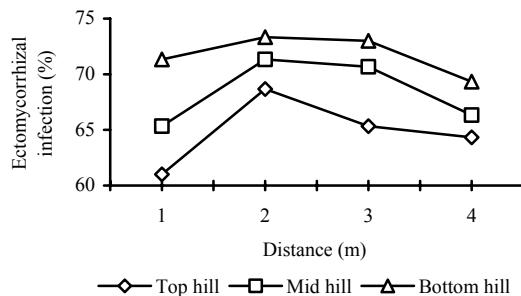


Fig. 2 ECM infection (%) in soil at 1, 2, 3 and 4m distances from the tree at three hill positions of 10-year-old *D. turbinatus* plantation

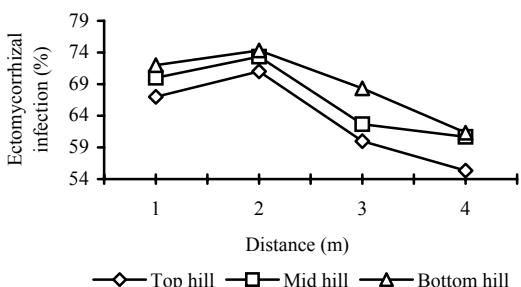


Fig. 3 ECM infection (%) in soil at 1, 2, 3 and 4m distances from the tree at three hill positions of 5-year-old *D. turbinatus* plantation.

The comparisons of ECM infection (%) in 5, 10 & 20-year-old plantations at top, mid and bottom hill positions are presented in Figures 4, 5 and 6, respectively. According to the age of plantations, the comparison of infection at different hill elevations varied considerably. At the top hill position the maximum infection was 71.00% found in 4 m distance in 20-year-old plantation compared with 10 and 5-year-old plantations (Figure 4). In the middle of the hill, the highest infection was 78.33% in 4 m distance from the base of the tree in 20-year-old plantation and the lowest was 60.67% in 1m distance in 5-year-old plantation (Figure 5). In case of bottom hill position, the infection was found highest in 20-year-old plantation followed by 10-year and 5-year old plantations (Figure 6).

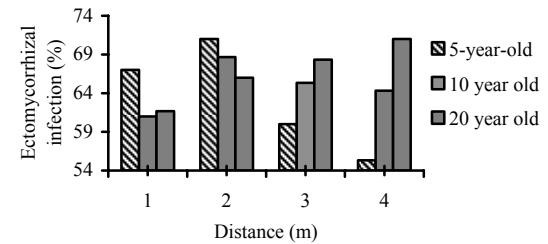


Fig. 4 Comparison of ECM infection (%) in soil at different distances from the tree at top hill position in 5, 10 and 20-year-old *D. turbinatus* plantations

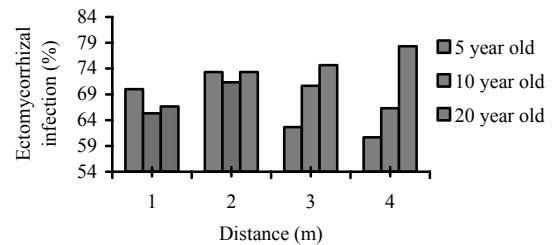


Fig. 5 Comparison of ECM infection (%) in soil at different distances from the tree at mid hill position in 5, 10 and 20-year-old *D. turbinatus* plantations

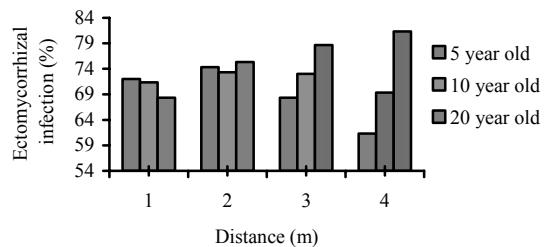


Fig. 6 Comparison of ECM infection (%) in soil at different distances from the tree at bottom hill position in 5, 10 and 20-year-old *D. turbinatus* plantations

Discussion

Most of the ECM roots were found in 1m and 2m distances from the trees in 5-year and 10-year-old plantations. With increasing distances, the percentages declined rapidly. In 20-year-old trees, the ECM infection (%) sharply increased with increasing distances from the base of the trees in three hill positions. Such a distribution ECM roots might be a general tendency

in natural mature forests, and it might be related to the existence of high amounts of nutrients that are available for plant roots and fungi. 95.4%, 83.5%, 76.4%, and 86.4% of root tips were found ectomycorrhizal in a survey of *Shorea leprosula* and *S. curtisii* in an underlogged and logged forest in Malaysia (Lee and Lim, 1987). The tendency of fine roots and ECM roots being mainly at the soil surface was found in various natural mature forests, including both broad-leaves and conifers (Büttner and Leuschner, 1994). The similar results were also found by Kimmins and Hawkes (1978), Vogt *et al.*, (1981), Ehrenfeld *et al.*, (1992) and Hashimoto and Hyakumachi (1998). Alexander *et al.*, (1992) reported that the ECM infection (%) declined markedly in distances greater than 30 meter from *D. costulata* tree. The proportion of living roots with ECMs remained high, at about 83%, above 30 cm depth in a *Betula pubescens* and *B. pendula* stand (Ingleby *et al.*, 1985). The percentage of ECM infection was found highest in mid hill position in 20-year-old pine trees at CUC in Bangladesh (Rahman and Mridha, 2004). Their result, showing a high concentration of ECM roots at the surface of the soil, was the same with present study.

Mycorrhization of forest crops has attracted considerable attention over the last few years because of their role as biofertilizers (Mridha, 2002), improving host growth as well as contributing to disease suppression (Marx, 1972). Hence, the appropriate association of ECMs with *D. turbinatus* is of considerable significance and needs further study and exploitation.

References

Alexander, I.J., Ahmad, N. and Lee, S.S. 1992. The role of mycorrhizas in the regeneration of some Malaysian forest trees [J]. Philippine Transaction Research Society, **335**: 379–388.

Alexander, I.J. and Hogberg, P. 1986. Ectomycorrhizas of tropical angiospermous trees [J]. New Phytologist, **102**: 541–549.

Aniwat, C. 1987. Mycorrhizal survey of dry-deciduous and semi-evergreen dipterocarp forest ecosystems in Thailand [C]. In: Kostermans, A. J. C. H. (ed.), *Proceedings of the Third Round Table Conference on Dipterocarps*. Mulawarman University, Indonesia, pp. 81–103.

Anon, 1989. Forest Management Plan. Management Plan for Chittagong University Campus for the period 1989–1990 to 1999–2000, University of Chittagong, Bangladesh.

Anon, 1979. Detailed soil survey of Chittagong University Campus, Chittagong, Department of soil survey, Government of the People's Republic of Bangladesh. 207pp.

Ashton, P.S. 1982. Dipterocarpaceae [J]. Flora Malaysian, Series, **19**: 237–552.

Bakshi, B.K. 1974. Mycorrhiza and its role in forestry [R]. Forest Research Institute, Dehra Dun, India.

Becker, P. 1983. Ectomycorrhizas on *Shorea* (Dipterocarpaceae) seedlings in lowland Malaysian rainforest [J]. Malaysian Forester, **46**: 146–170.

Buttner, V. and Leuschner, C. 1994. Spatial and temporal patterns of fine root abundance in a mixed oak-birch forest [C]. Forest Ecology Management, **70**: 11–21.

Das, S. 1980. Dipterocarp forests of Bangladesh and their management [J]. Bano Bigyan Patrika, **8**: 71–86.

de Alwis, D.P. and Abeynake, K. 1980. A survey of mycorrhizas in some tropical forest trees of Sri Lanka [C]. In: Mikola, P. (ed.), *Tropical mycorrhiza research*. Oxford: Clarendon Press, pp. 146–153.

Ehrenfeld, J.G., Kaldor, E. and Parmelee, R.W. 1992. Vertical distribution of roots along a soil toposequence in the new Jersey Pinelands [J]. Can. J. For. Res., **22**: 1929–1936.

Hashimoto, Y. and Hyakumachi, M. 1998. Distribution of ectomycorrhizas and ectomycorrhizal fungal inoculum with soil depth in a birch forest [J]. Journal of Forest Research, **3**: 243–245.

Hadi, S. and Santoso, E. 1988. Effect of *Russula* spp., *Scleroderma* sp. and *Boletus* sp. on the mycorrhizal development and on the growth of five dipterocarp species [J]. In: Singh, M. M. (ed.), *Agricultural and biological research priorities in Asia*. International Foundation for Science and Malaysian Scientific Association, pp. 183–185.

Hadi, S., Fakuara, Y., Setiadi, Y., Prematuri, R. and Nuhamara, S.T. 1991. Status of mycorrhiza research on Dipterocarps in Indonesia. In: *Proceedings of BIO-REFOR Pre-Workshop*, Bogor, Indonesia, pp. 75–81.

Hong, L.T. 1979. A note on dipterocarp mycorrhizal fungi [J]. Malaysian Forester, **42**: 280–283.

Ingleby, K., Last, F.T. and Mason, P.A. 1985. Vertical distribution and temperature relations of sheathing mycorrhizae of *Betula* spp. growing on coal spoil [J]. Forest Ecology Management, **12**: 279–285.

Ingleby, K., Mason, P.A., Last, F.T. and Fleming, I.V. 1990. Identification of ectomycorrhizas Institute of Terrestrial Ecology[R]. Research publication No. **5**, 112 pp.

Kimmins, J.P. and Hawkes, B.C. 1978. Distribution and chemistry of fine roots in white spruce sspalpine fir stand in British Columbia: implication for management [J]. Can. J. For. Res., **8**: 265–279.

Lee, S.S. 1998. Root Symbiosis and Nutrition [C]. In: Appanah, S. and Turnbull, J. M. (eds.), *A Review of Dipterocarps: Taxonomy, Ecology and Silviculture*. Center for International Forestry Research. Forest Research Institute Malaysia, pp. 98–114.

Lee, S.S. and Lim, K.L. 1989. Mycorrhizal infection and foliar phosphorus content of seedlings of three Dipterocarps species growing in a selectively logged forest and a forest plantation [J]. Plant and Soil, **117**: 237–241.

Marx, D.H. 1972. Ectomycorrhizae as biological deterrents to pathogenic root infections [J]. Annual Review of Phytopathology, **10**: 429–434.

Marx, D.H. 1977. The role of mycorrhizae in forest production [J]. Tappi, **60**: 151–161.

Mridha, M.A.U. 2002. The Potential application of arbuscular mycorrhizal fungi in hill farming systems in the CHT [C]. In: Khan, N. A.; Alam, M. K.; Khisa, S. K. and Millat-e-Mustafa, M. (eds.), *Farming Practices and Sustainable Development in the Chittagong Hill Tracts*. CHTDB and VFFP-IC. Bangladesh, pp. 237–246.

Rahman, M.S. and Mridha, M.A.U. 2004. Status of ectomycorrhizal association in *Pinus caribaea* grown in Chittagong University Campus [J]. Journal of Forestry and Environment, **2**: 67–70.

Rawat, P.S., Ginwal, H.S., Singh, R.P. and Duvey, R.C. 2003. Vertical distribution of ectomycorrhizae in deodar and chir pine forests in relation to their soil characteristics [J]. The Indian Forester, **129**(5): 624–630.

Shayesta, B. and Choudhury, J.A. 1985. Effect of inoculation of mycorrhizal fungi on the growth and survival of *Pinus oocarpa* in the nursery [J]. Bangladesh Journal of Agriculture, **10**: 47–52.

Singh, K.G. 1966. Ectotrophic mycorrhiza in equatorial rain forests [J]. Malaysian Forester, **29**: 13–18.

Smits, W.T.M. 1987. Production of Dipterocarps planting stock in nurseries [C]. Pp. 151–157. In: Kostermans, A. J. C. H. (ed.), *Proceedings of the Third Round Table Conference on Dipterocarps*. 16–20 April, 1985, Samaria, Indonesia, UNESCO, Indonesia.

Vogt, K.A., Edmons, R.L., Grier, C.C. 1981. Seasonal changes in biomass and vertical distribution of mycorrhizal and fibrous-textured conifer fine roots in 23- and 180-year old subalpine *Abies amabilis* stands [J]. Can. J. For. Res., **11**: 223–229.

Yasman, I. 1993. Ectomycorrhizal sporocarp appearance in a dipterocarp forest, East Kalimantan, Indonesia [C]. In: Suzuki, K., Sakurai, S. and Ishii, K. (eds.), *Proceedings of the International Workshop of BIO-REFOR.. 20-23 September 1993, Yogyakarta, Indonesia*, BIO-REFOR/IUFRO/SPDC, Tsukuba, Japan, pp. 179–181.

Zarate, J.T., Watling, R., Jeffries, P., Dodd, J.C., Pampolina, N.M., Sims, K., Lorilla, E.B. and de la Cruz, R. 1993. Survey of ectomycorrhizal fungi associated with pines and dipterocarps in the Philippines [C]. In: Suzuki, K., Sakurai, S. and Ishii, K. (eds.), *Proceedings of the International Workshop of BIO-REFOR.. 20-23 September 1993, Yogyakarta, Indonesia*, BIO-REFOR/IUFRO/SPDC, Tsukuba, Japan, pp. 182–185.